OTES

an using polyamide layers. The content (16%) of polyamide in this polyamide-silica el mixed layer is higher than that in the previous report $(12\%)^4$ for getting a more able layer.

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J. Chromatog., 44 (1969) 201–203

HROM. 4284

olyamide-kieselguhr thin-layer chromatography of yellow food dyes

The thin-layer chromatography of food dyes has been studied by numerous nvestigators. The separation of synthetic food dyes by thin layers of cellulose¹, tarch², silica gel³, aluminum oxide⁴ and polyamide⁵ has been reported. Recently, a etter separation of red food dyes was obtained by CHIANG⁶ with a mixed polyamideilica gel thin layer. Therefore, further modification of this method was tried. In this ote, the separation of five yellow food dyes and three harmful yellow dyes (auramine, netanil yellow and picric acid) by mixed polyamide-kieselguhr thin-layer chromato-raphy is described. For comparison, the thin-layer chromatography on only poly-mide and on only kieselguhr is also described.

Experimental

Preparation of polyamide-kieselguhr mixed layer. Ten grams of polyamide chip Nylon 6, type 1022B of UBE Industrial Ltd., Osaka, Japan) were dissolved in 80 ml f 90% formic acid; then 20 ml of distilled water were added. After warming (below 0°) and stirring, a homogeneous solution was obtained. It was then cooled to room emperature, and 40 grams of Kieselguhr G (E. Merck) were added. Of the previous olution 200 ml were poured into a dish $(14.5 \times 19.5 \times 2.5 \text{ cm})$ into which a glass plate $12 \times 14 \times 0.1 \text{ cm}$) was dipped. Both sides of the glass were covered homogeneously. The glass was hung for 2 min over the dish to let the excess solution drain off. It was hen air dried for 3 h and heated at 100° for 30 min.

Preparation of polyamide layer. Instead of 10 g, 20 g of polyamide were dissolved before proceeding as described in the previous method, but without adding Kieselguhr G.

Preparation of kieselguhr layer. Dilute slurries of Kieselguhr G (45 g in 100 ml of water) were sprayed at 2 kg/cm³ pressure from a distance of 20 cm onto 8 horizontal glass plates (12×14 cm) which were then dried at 100° for 30 min. The thickness of the ayers was about 250 μ .

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TABLE I

CHROMATOGRAPHIC DATA

Solvent I: methanol-acetone-water-30% sodium acetate solution-ethylenediamine (10:10:20:5:2); solvent II: ethanol-water-ether-5% NH₄Cl solution-ethylenediamine (15:15:10:5:2). a, R_F value obtained on polyamide-kieselguhr layer; b, kieselguhr layer; c, polyamide layer.

No.	Dyes	Solvent I			Solvent II		
		a	ь	С	a	ь	C
I	Naphthol yellow S	0.66	0.97	0.45	0.69	0.98	0.54
2	Yellow AB	0.11	0.82	0.02	0.35	0.88	0.10
3	Yellow OB	0.05	0.79 ^ª	0.01	0.23	0.84	0.10
4	Tartrazine	0.91	0.97	0.84	o.88	0.85	0.80
5	Sunset yellow FCF	0.38	0.98	0.59	0.81	0.94	0.78
6	Metanil yellow	0.31	0.88	0.13	0.53	0.92	0.43
7	Auramine	0.73	0.96	0.35 ^a	0.76	0.96	0.49
8	Picric acid	0.53	0.98	0.77 ^a	0.60	0.95	0.52
Time required (min) ^b		75	30	300	150	90	600

^a Tailing.

^b Time required to ascend 10 cm from origin.

Chromatographic procedure. A 0.5% alcoholic solution of Yellow AB, Yellow OB and auramine and a 0.5% water solution of other dyes were applied to the starting line 1.5 cm from the bottom of the layer, and the plate was developed by ascending techniques. The chamber had been equilibrated with the respective solvent for 30 min before use.

Results and discussion

The R_F values obtained with two solvent systems are given in Table I. It has been found that the results show better separation and sharp spots with polyamidekieselguhr mixed layers than with polyamide and kieselguhr layers. Also a 10-cm ascent from the origin is more rapid using the mixed layers than when polyamide layers are employed. In the mixed layer, polyamide serves as a strong binder and makes the layers very durable and easy to handle. Both sides of the glass are independent of each other, and chromatography can be performed simultaneously on both sides. The addition of a small amount of salt and ethylenediamine in the solvent mixture is essential to break hydrogen bonding between polyamide and dyes. Oil-soluble dyes of Yellow AB and Yellow OB are rather difficult to separate because of the close similarity of their structures.

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First received June 9th, 1969; revised manuscript received July 11th, 1969

J. Chromatog., 44 (1969) 203-204

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